

Novel, heterocyclic small molecule inhibitors of PD-1/PD-L1 pathway

Shivani Garapaty, Dhanalakshmi Sivanandhan, Chandregowda Venkateshappa, Guru Pavan Kumar Seerapu, Reshma Das, Pradeep Nagaraj, Ronodip Kar, Anuj Kumar Singh, Venkatesha Ashokakumar Venkatasubbaiah, Rama Kishore VP Putta, Muralidhar Pendyala, Girisha Lokesh, Hari Madaka, Harikrishna Reddy Thummuru, Shikas AP, Pratheeksha Anchan, Prathima Bhat, Rudresha G., Mohd. Zainuddin, Krishnakumar V, Ramachandraiah Gosu, Rajendra Kristam, Jeyaraj DA, Sriram Rajagopal. Jubilant Biosys Ltd., Bangalore India

SUMMARY

The PD-1/PD-L1 molecular pathway is one of the primary mechanisms of immune evasion deployed by cancer cells. Induction of PD-L1 expression on cancer cells is associated with inhibition of immune responses against cancer, thus permitting cancer progression and metastasis. Activation of PD-1/PD-L1 pathway induces apoptosis of activated T-cells, inhibits their proliferation, facilitates T-cell anergy and exhaustion and enhances the function of regulator T-cells. Therefore, blocking this pathway restores the proliferation and cytotoxicity of CTLs, inhibits the function of Tregs and results in decreased T-cell apoptosis. Although a number of therapeutic antibodies targeting PD-1/PD-L1 have been developed and approved for a number of malignancies, there is a still a need for potent, selective small molecule inhibitors of the PD-1/PD-L1 pathway

Rational and structure guided *de novo* design approaches were used to design novel small molecule PD-1/PD-L1 pathway inhibitors; potency of these inhibitors was assessed in an *in-vitro* TR-FRET assay. Checkpoints signaling reporter assays as well as ex-vivo co-culture assays were used to assess the ability of the compounds to restore T-cell proliferation and function.

Three novel chemical series as potent PD-1/PD-L1 pathway inhibitors are being developed for the treatment of cancer. A representative inhibitor JBI-426 showcased here exhibited low nM potency in vitro and no cytotoxicity against cancer cell proliferation per se. JBI-426 showed good in vitro ADME properties in terms of aqueous solubility, metabolic stability, permeability and excellent oral bioavailability in mouse pharmacokinetics. In a RENCA syngeneic model, oral administration of JBI-426 at 50 mg/kg resulted in a strong tumor growth inhibition, comparable (or better) than the PD-L1 mAb, and was well tolerated. The effect of JBI-426 on tumor infiltrating lymphocytes was also assessed; a significant increase in CD8+ cytotoxic lymphocytes was observed. The anti-tumor effect of JBI-426 was also exhibited in CT-26 syngeneic model.

Biochemical characterization Two representative compounds Series 2 Series 3 Series 1 0.048 0.066 0.022 IC₅₀ 0.043 0.083 0.039 Series 1 Series 2 Series 3 0.0010.01 0.1 1 10 100 0.0010.01 0.1 1 10 100 0.0010.01 0.1 1 10 100 compound [µM] compound [µM] compound [µM] IC50 0.02182 IC50 0.06573 IC50 0.04802







Top: Purified T-cells were isolated from 6-8wk C57BL/6 female mice and co-cultured with either Recombinant PD-L1 protein or RENCA cells. Secreted IFNy levels was assessed by ELISA. Bottom: PD-L1 is endogenously expressed in RENCA cells; JBI-426 does not affect the viability of PBMCs, murine T-cells or RENCA

Pharmacokinetic profile of JBI-426



Contact: dhanalakshmi.sivanandhan@jubilantbiosys.com

rsus Time	Parameter, PO	Mice	
alb/c mice	t _{1/2} (hr)	2.07	
	Clearance (ml/min/kg)	3.98	
	C _{max} (ng/ml)	7395	
	AUC _(0-t) (ng*hr/ml)	29715	
	AUC _(0-∞) (ng*hr/ml)	29731	
20 24	t _{max} (hr)	0.25	
	t _{last} (hr)	24	

Bright CD4 CD3 SSC field CD8

PBMCs were isolated from a healthy volunteer and co-cultured with A375 cells, treated with IFNy. Post compound treatment for 48H, PBMCs were stained with CD3-PE, CD4-FITC and CD8-APC to assess the T-cell population. Image-based FACS acquisition on Amnis® Flowsight



Gene	PBMCs vs. CC	CC vs. JBI-426	Gene	PBMCs vs. CC	CC vs. JBI-426	Upregulated Pathways	FDR
PMCH	-4.47	4.59	IL1RL1	9.49	-7.87	Natural killer cell	
IFNG	-4.3	1.75	IL13RA2	89.66	-7.6	mediated cytotoxicity	5.16e-02
GZMA	-4.14	2.53	CT45A1	117.66	-5.55		4 9 9 9
IL17F	-3.47	8.09	AXL	69.78	-5.44	I cell activation	1.8e-01
LRRN3	-3.25	2.08	APOE	4.71	-4.87	Jak-STAT signaling	8.65e-01
BIRC5	-3.23	2.19	MAGEA3	28.3	-4.77	pathway	
CDK1	-2.87	2.06	THBS1	13.43	-4.52	p all may	
TTK	-2.84	2.09	C1S	24.6	-4.23	Downregulated	
DPP4	-2.84	2.65	TLR4	4.46	-4.13	Downlegulated	FDR
GZMB	-2.8	3.76	TLR8	5.64	-3.82	Patnways	
IL32	-2.75	1.96	IL24	174.05	-3.79	Chemokine signaling	1 570 01
ICOS	-2.57	1.75	C2	7.03	-3.7	pathway	1.576-01
GZMH	-2.46	1.81	FCGR2A	3.94	-3.7	PI3K-Akt signaling	
TUBB	-2.42	1.5	LRP1	5.39	-3.68	nathway	5.87e-01
PBK	-2.34	1.81	CLEC5A	4.46	-3.68		
HAVCR2	-2.18	2.38	SPANXB1	18.17	-3.67	CC: Co-culture	
CD8A	-2.15	1.73	CYBB	6.7	-3.63	nanoString	
CD3G	-2.09	1.53	CD276	8.73	-3.48	TECHNOLOGIES	
LTB	-1.99	1.92	APP	8.77	-3.34	theracues	
CD8B	-1.97	1.73	LILRB3	5.48	-3.33	SIGNALS FOR LIFE	

KEGG pathway analysis of differentially expressed genes in PBMCs cocultured with A375, in the presence or absence of JBI-426



PD-L1 mAb



JBI-426 is efficacious in CT-26 syngeneic model

TILs were assessed in the two treatment groups: JBI-426 administration

resulted in a more significant increase in CD8+ TILs as compared to anti-

(0.1mg/mouse, Q4D) in

RENCA syngeneic model.



Conclusions

Small molecule PD-1/PD-L1 inhibitors, in contrast to antibody therapies, can provide increased oral bioavailability, increased bioefficiency and shorted half life activity for a more controllable treatment, particular in the case of auto-immune or other adverse effects.

Further studies to assess additional compounds from the three chemical series are underway. The oral administration route of these PD-1/PD-L1 inhibitors would provide an attractive alternate to the currently available antibodies in treating cancer either as a stand-alone therapy or in combination with other immuno-modulatory agents, as well as other standard of care agents.